RESEARCH ARTICLE

Resveratrol and prostate cancer: Promising role for microRNAs

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Scope: Resveratrol (Res) has anticancer activity in prostate cancer (PCa), which can be attributed to modulation of microRNAs (miRNAs/miRs). miRNAs/miRs are small noncoding RNAs that negatively regulate gene expression. We have analyzed differential miRNA expression in PCa cells treated with Res.

Methods and results: Using miRNA microarrays we found that 23 miRNAs were significantly down-regulated and 28 miRNAs were significantly up-regulated after Res treatment. The down-regulated miRs included miR-17-92 and miR-106ab clusters with well recognized oncogenic properties while the up-regulated miRs included several tumor suppressors. Selected miRs were verified by qRT-PCR, including miR-17, miR-20a, miR-20b, miR-106a and miR106b. Since these miRNAs target PTEN (phosphatase and tensin homolog deleted on chromosome 10), we performed Western blot to confirm up-regulation of PTEN in PCa cells. In addition, using TargetScan database, we have identified putative mRNA targets for Resinduced down- and up-regulated miRs. Using a bioinformatics approach, we generated gene networks specifically altered by Res-regulated miRNAs.

Conclusion: Our results indicate that the dietary compound Res may play an important role in prostate carcinogenesis through modulation of miRNA expression: Res down-regulated oncogenic miRs and up-regulated tumor suppressor miRs in PCa cells. Further in-depth studies are necessary in order to fully recognize the beneficial miRNA-mediated effects of Res in PCa.

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Keywords:

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1 Introduction

Prostate cancer (PCa) is the most common male malignancy and the second cause of cancer-related death in the US.

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Abbreviations: AR, androgen receptor; GO, gene ontology; miRNA/miR, microRNA; LNCaP, Lymph node cancer of prostate; MTA1, metastasis-associated antigen 1; oncomiR, oncogenic miRNA; Par-4, prostate apoptosis response protein 4; PCa, prostate cancer; PSA, prostate specific antigen; PTEN, phosphatase and tensin homolog deleted on chromosome 10; Res, resveratrol; RPMI, Roswell Park Memorial Institute; SIRT1, sirtuin 1; SOCS, suppressor of cytokine signaling; SOX, SRY related HMG box; UTR, unfranslated region

Nutritional chemoprevention is a particularly promising approach for PCa because of its slow progression and predominance in elderly men. Resveratrol (Res) has been proposed as an especially suitable agent for PCa due to its specific androgen receptor (AR)-related effects [1]. In addition, epidemiological data indicate a reduced PCa risk associated with red wine consumption, attributable mostly to high Res content [2].

Res (3, 5, 4'trihydroxy-trans-stilbene), a natural phytoalexin, is known for its cardioprotective, anti-inflammatory and anticancer activities [3]. Res has pleiotropic anticancer activities that include induction of cell cycle arrest and apoptosis [4–6] and inhibition of angiogenesis, invasion and metastasis [7–10]. Little is known on the epigenetic mechanisms of Res chemoprevention and anticancer effects. Accumulated data suggest that beneficial effects of dietary polyphenols may be in part attributed to their epigenetic properties, including changes in DNA

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methylation, regulation of histone modifications and changes in the expression of microRNAs (miRNAs/miRs) [11]. However, this is an area that is mostly underexplored, particularly in PCa [12]. Currently, epigenetic mechanisms of Res are associated with the ability of Res to activate histone acetyl transferase (HAT) p300 and histone deacetylase sirtuin 1 (SIRT1) [13, 14]. The exact mechanism by which Res activates SIRT1 remains controversial [15, 16]. The apparent paradox that Res can simultaneously activate histone acetyl transferases and a deacetylase remains unexplained. In contrast, we recently demonstrated that Res acts as natural indirect histone deacetylase (HDAC) inhibitor by reversing the activity of co-repressor nucleosome remodeling and deacetylation complex (NuRD) involved in histone deacetylation and gene-specific transcriptional regulation: Res down-regulates metastasis-associated antigen 1 (MTA1), a protein that has pro-survival, anti-apoptotic, invasive and pro-angiogenic properties in PCa [17, 18]. We further demonstrated that MTA1 silencing through RNA interference significantly sensitized the PCa cells to Resdependent p53 acetylation and apoptosis [19].

Another important epigenetic network of gene regulation is represented by miRNAs [11, 20]. Recently, few reports indicated that dietary factors play an important role in the process of carcinogenesis through modulation of miRNA expression [21, 22]. Therefore, modulation of miRNA expression may contribute to the cancer-protective effects of Res known as a potential chemopreventive and therapeutic agent in PCa. miRNAs are small (~18-22 nucleotides), non-coding, single-stranded, sequence-specific RNAs that negatively regulate gene expression by binding complementary sequences in the 3' untranslated regions (UTRs) of target mRNA and inhibiting stability and/or translation of mRNA [23, 24]. miRNAs act as oncogenes or tumor suppressors in cancer [25]. Oncogenic miRNAs (oncomiRs) that are overexpressed in different cancers usually promote tumor development by inhibiting tumor suppressor genes and genes that control cell differentiation and apoptosis. In contrast, tumor suppressor miRNAs have decreased expression in cancers and usually prevent tumor development by negatively affecting oncogenes and promoting genes that control cell differentiation and apoptosis. Various miRNA expression profiling studies reported on both upand down-regulation of miRNAs in PCa tissues compared to normal prostate tissue or benign peripheral zone tissue from radical prostatectomy specimens [26, 28]. Some changes in miRNAs have been correlated with Gleason score and clinical recurrence [29, 30]. Thus, differential miRNAs in PCa can be useful diagnostic and prognostic indicators. Because of high enthusiasm about the use of Res in human chemoprevention [31, 32], the ability of Res to alter miRNAs offers new perspectives for Res chemoprevention and anticancer activities. However, there are no reports on Resdependent alterations of miRNA expression in PCa.

To our knowledge, this is the first study to look at changes in miRNA levels by Res in PCa. In this study, we

showed that Res significantly down-regulated several miRNA oncogenic clusters, up-regulated some known tumor suppressors and modulated other miRNAs whose biological role has yet to be established.

2 Materials and methods

2.1 Materials

Res was purchased from Sigma (St. Louis, MO, USA). Stock solution of Res was made using 100% ethanol and kept at 4°C, in the dark. Lymph node cancer of prostate (LNCaP) and Du145 cells were maintained in RPMI medium containing 10% FCS as described previously [18, 19]. Cells were maintained in phenol red-free media with 5% charcoal-stripped calf serum overnight prior to Res treatment.

2.2 Array-based miRNA expression profiling and analysis

LNCaP cells were maintained in RPMI 1640 medium containing 10% FCS as we previously described [18, 19]. When cells achieved about 60% confluence, the media was replaced with phenol red-free RPMI supplemented with 5% charcoal-stripped calf serum overnight to eliminated hormonal background that might interfere with Res effects. Res or EtOH-vehicle was added to the media for the next 24 h after which RNA was isolated using miRNeasy Mini Kit (Qiagen, Valencia, CA). We used miRNA microarray detection service from LC Sciences (Houston, TX, USA) using MRA 1001 chip, which contains 894 human miRNA probes listed in Sanger miRBase Release 14.0 (http:// www.sanger.ac.uk/Software/Rfam/mirna/). RNA integrity and purity was checked before analysis. The miRNA-enriched samples were then labeled with fluorescent dyes for two color detection (Cy3 and Cy5). Hybridization, washing and detection of miRNA array chips were controlled to optimize signal. After background subtraction and normalization, data analysis was performed, and a list of up- and down- regulated transcripts with statistical significance of the *p*-values < 0.01 was provided.

2.3 Identification of miRNA putative target genes and network analysis

TargetScan database, version 5.1 (www.targetscan.org) was used for identification of putative targets for identified miRNAs. This algorithm predicts miRNA targets on the basis of sequence complementarity and thermostability [33]. The overall target specificity is expressed by a context score [34] that combines the complementarity of seed sequence, site position and local adenine/uridine (AU) content to evaluate site efficacy between miRNAs and mRNA targets.

To enhance the accuracy for miRNA target prediction and select high confidence targets, only genes with total context score $\leq\!-0.5$ were included in our analysis. We generated regulatory networks and visualization using Bayesian modeling for the putative targets of two co-regulatory miRNAs sets: oncogenic and tumor suppressor. We then integrated these genes onto a global molecular network (gene ontology (GO) database), which identifies the biological functions that are most significant to the genes in the network.

2.4 Validation of miRNA expression by real-time RT-PCR

RNA was isolated using miRNeasy Mini Kit (Qiagen) according to the manufacturer's instructions. qRT-PCR was performed using mercury LNATM Universal RT miRNA PCR (Exigon, Woburn, MA). After first-strand cDNA synthesis, the real-time amplification was carried out using cDNA template (8 µL), SYBR Green master mix (10 µL) and miRNA specific primer mix (2 µL) in a final volume of 20 µL. The condition for PCR was 95°C for 10 min, followed by 40 cycles of 95°C for 10 s and 60°C for 1 min, performed in a CFX96 Real-Time System (Bio-Rad). All primers used for qRT-PCR were purchased from Exigon. SNORD44 was used as an internal reference standard. Fold change of normalized miRNA was calculated using the $2^{-\Delta\Delta Ct}$ for each miRNA by double-delta C_t method [35]. Student's two-tailed paired t-test was performed to analyze results from qRT-PCR.

2.5 Western blot

Western blot analysis was performed as described previously [18, 19, 36]. Briefly, the cells were maintained in regular RPMI 1640 media, and then media was changed to phenol red-free supplemented with stripped serum 24 h before treatment with Res. Cells were harvested using lysis buffer. Equal amounts of protein were size-fractionated by 7.5% Tris-HCl Ready gels (Bio-Rad Labs, Hercules, CA) and transferred to polyvinylidene fluoride (PVDF) membrane. The membranes were then blocked with Tris buffered saline (TBS)-Tween and 5% dry milk and subsequently probed with anti-PTEN (PTEN, phosphatase and tensin homolog deleted on chromosome 10) and anti-β-actin antibodies (Santa Cruz, Santa Cruz, CA). Signals were visualized using enhanced chemiluminescence. Images were quantified using Image J software.

3 Results and discussion

3.1 Res alters miRNA expression in human PCa cells

Res induces apoptosis, inhibits cancer cell growth, angiogenesis and metastatic potential of PCa cells in vitro and in vivo [19, 37-40]. These effects of Res could be mediated via the regulation of miRNAs. As mentioned, studies reported on both up- and down-regulation of miRNAs in PCa [26-28]. Moreover, the link between miRNAs changes and the development and progression of PCa has been established [41-43]. Since Res is an anticancer agent in PCa, Resinduced down-regulated miRNAs must function as oncomiRs whereas Res-induced up-regulated miRNAs must function as tumor suppressors. We performed miRNA microarray analysis of Res-treated LNCaP cells and found that 51 miRNAs were modulated, from which 23 miRs were significantly down-regulated (oncomiRs) and 28 miRs were significantly up-regulated (tumor suppressor miRNAs) following treatment with 50 µM Res (Fig. 1). Each group of statistically significant miRNAs is represented as two subgroups: above the horizontal line are transcripts with high signal intensity and below are transcripts with low signal intensity (Fig. 1B). We are interested in the differences between expression levels for a given miRNA in untreated and Res-treated cells independent of its basal levels of expression. In fact, for most miRNAs with low signals (low abundance), the changes in expression after Res treatment were more dramatic than for miRNAs with higher abundance. These changes could be of particular interest and should not be overlooked.

We performed a vigorous literature and web search to draw a parallel between our findings and already known roles of miRNAs in human malignancies, particularly in PCa. First, we generated two lists of potential gene targets for identified onco- and tumor suppressor miRNAs using TargetScan database (http://www.targetscan.org), which predicts target mRNAs by seed sequence complementarity (Supporting Information data, ST1 and ST2). To select high confidence targets we used single criterion of the context score threshold and filtered out targets with score >-0.5. The total number of putative targets predicted for Resregulated oncogenic and tumor suppressor miRNAs are shown in Table 1. We then compared our data to the miRNA expression reported by others, in which miRNAs were classified as oncogenes or tumor suppressors. We first searched for oncomiRs. Remarkably, Res significantly down-regulated five (miR-17, miR-18b, miR-20a, miR-20b and miR-92b) from six members of the c-MYC-activated oncomiR-17-92 cluster [44]. This cluster is amplified in lung cancer and lymphoma, frequently overexpressed in various cancers, including PCa, and known to accelerate angiogenesis and promote tumor formation in mouse models [25, 26, 45-48]. As an example, overexpression of miR-20a inhibited apoptosis in PCa cell lines and inhibition of miR-20a resulted in increased apoptosis after doxorubicin treatment [49]. Overexpression of cell cycle transcription factor E2F1, which is one of the targets of the miR-17-92 cluster, induced apoptosis in LNCaP cells [47, 50]. Further, Res inhibited miR-106a and miR-106b oncomiRs that are reported to be highly expressed in PCa and mediate antiapoptotic activity by targeting p21/WAF1 and E2F1 [41]. Notably, both

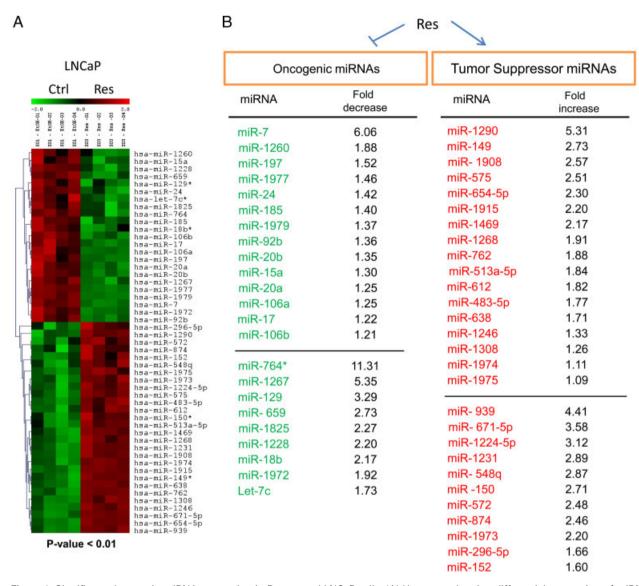


Figure 1. Significant changes in miRNA expression in Res-treated LNCaP cells. (A) Heat map showing differential expression of miRNAs (hsa-miRs) in LNCaP treated with either EtOH (control, trl) or $50\,\mu\text{M}$ Res. p < 0.01. The relative transcript abundance of each miR is color coded: green indicates low expression; black indicates intermediate and red indicates high expression. (B) Results of analysis ranked by fold change. A total of 23 miRs (oncomiRs) were significantly down-regulated (green) and 28 miRs (tumor suppressor miRNAs) were significantly up-regulated (red). Fold decrease/increase was calculated from $\log_2 \text{Res/Ctrl}$ values. Horizontal lines in each set separate two subgroups of miRNAs: transcripts with higher abundance (above the line) and transcripts with low abundance (below the line). Note: hsamiR hereafter referred to as miR.

miR-17-92 and miR-106b-25 clusters are regulators of a critical tumor suppressor gene PTEN, and miRs from these clusters directly associate with PCa tumorigenesis [51]. For example, the miR-106b-25 locus in chromosome 7 is over-expressed and amplified in PCa. Moreover, miR-106a-25 expression is inversely correlated with PTEN abundance in human PCa tissues [51]. Since PTEN is frequently defective in PCa as its deletions/mutations are found in 30% of primary and in 63% of metastatic disease [52, 53], Res rescue of PTEN by down-regulation of the oncogenic PTEN-targeted miRNA-network is of considerable interest.

Among the most significantly down-regulated miRNAs by Res were miR-7 (6.06-fold decrease) and miRs from a second subgroup of oncomiRs with a fold decrease ranging from 1.92 to 11.31 (Fig. 1B). The changes of low abundant miRNAs after Res treatment were more pronounced than in miRNAs with high signals and might have more biological significance. However, little is known about the cancer-related functions of most of the identified miRs in this subgroup. High expression of miR-1228 was reported in malignant mesothelioma compared to normal samples [54]. Expression of miR-129 (and corresponding low SRY-related HMG box

Table 1. Res-regulated miRNAs and numbers of their putative targets

Oncogenic	Targets	Tumor suppressor	Targets
miRNAs	(<i>n</i>) ^{a)}	miRNAs	(<i>n</i>) ^{a)}
miR-7	33	miR-1290	46
miR-1260	21	miR-149	17
miR-197	10	miR-1908	0
miR-1977	0	miR-575	4
miR-24	36	miR-654-5p	3
miR-185	34	miR-1915	0
miR-1979	0	miR-1469	0
miR-92b	0	miR-1268	1
miR-20b	3	miR-762	0
miR-15a	ა 10	miR-513a-5p	47
miR-20a		miR-612	47 5
miR-20a	21		0
	0	miR-483-5p	
miR-17	68	miR-638	0
miR-106b	3	miR-1246	14
miR-764	0	miR-1308	4
miR-1267	8	miR-1974	0
miR-129	0	miR-1975	0
miR-659	49	miR-939	3
miR-1825	6	miR-671-5p	10
miR-1228	4	miR-1224-5p	7
miR-18b	1	miR-1231	3
miR-1972	0	miR-548q	0
let-7c	1	miR-150	17
		miR-572	0
		miR-874	3
		miR-1973	0
		miR-296-5p	4
		miR-152	5

a) Target numbers are based on total context score ≤ -0.50 . Table is created using TargetScan database (www.target scan.org).

(SOX) 4 levels) in bladder cancer was found to be associated with poor outcome [55]. On the other hand, the expression of miR-129-2 was lost in primary endometrial tumors, and reactivation of miR-129-2 in cancer cells resulted in decreased SOX4 and reduced proliferation [56]. The biological functions of miR-764, miR-1267, miR-659, miR-1825 and miR-1972 have not been characterized in cancer. However, while miR-764 and miR-1972 showed no identified putative targets (TargetScan), miR-659, miR-1825 and miR-1267 indicated numerous targets that are implicated in cancer (Table 1). For example, putative targets of miR-659, suppressor of cytokine signaling (SOCS) protein family, play an important role in PCa: SOCS1 is expressed in PCa and acts as a negative growth regulator [57] whereas androgen-regulated SOCS2 seems to have a role in hormonal progression to androgenresistant disease [58].

Other down-regulated miRNAs by Res were miR-7, miR-197 and miR-24. The role of miR-7 in tumorigenesis is complex and undecided: it has been reported as both an oncomiR and tumor suppressor. Overexpression of miR-7 was documented in breast and lung cancer and was associated with poor outcome of diseases [59, 60]. On the other

hand, miR-7 has been characterized as a tumor suppressor targeting oncogenes and a number of key signaling molecules of survival pathways in several cancers [61-63]. It is likely that particular interaction between a miRNA and its target genes may be influenced by microenvironment and dependent on cancer type and cell specificity. Because Res down-regulated miR-7 in our study, we believe that miR-7 plays an oncogenic role in PCa. In agreement with this notion, prostate apoptosis response protein 4 (Par-4), a tumor suppressor protein that was isolated from PCa cells undergoing apoptosis, is linked to miR-7: in Par-4-transfected colon cancer cells, miR-7 was down-regulated among other oncomiRs (e.g. miR-18a) and the down-regulation was associated with up-regulation of apoptotic voltage-dependent anion channel protein 1 (VDAC1) [64]. Further studies are needed to elucidate the role of miR-7 in PCa. High levels of miR-197 were found in small-cell and non-small-cell lung carcinoma cell lines and in non-small-cell lung cancer tumor specimens, which correlated with reduced expression of target tumor suppressor Fus1 (equivalent of human TUSC2, tumor suppressor candidate 2) [65]. miR-24 is up-regulated in cervical and gastric cancers [26, 66]. Importantly, miR-24 has been suggested as a potential target for gene and drug therapy to treat hormone-insensitive PCa because the down-regulation of miR-24 induced apoptosis in Du145 cells via up-regulation of its target pro-apoptotic factor Fas-associated factor 1 (FAF1) [67]. To our surprise, Res down-regulated let-7c, miR-15a and miR-185 miRNAs, which have been reported as tumor suppressors [68–70]. No information was found on the roles of miR-1260, miR-1977, miR-1267, miR-1825. Altogether, Res widely down-regulated miRs with oncogenic properties that are overexpressed in various cancers, including PCa.

miRNAs up-regulated by Res fall into three major groups: (i) miRs that have been reported by others to be down-regulated in different cancers (miR-149, miR-152, miR-150; miR-939, miR-575, miR-483 and miR-654); (ii) miRs that are have been reported to be up-regulated (miR-572, miR-762 and miR-1246) and (iii) miRs whose aberrant expression or regulation have not been reported before and whose biological functions have not been characterized (miR-1290, miR-1908, miR-1915, miR-1469, miR-1268, miR-1231, miR-1973, miR-548q, miR-1974, miR-1975 and miR-671-5p). Some of the miRs from this group, however, have many putative target genes according to TargetScan database (Table 1). Although knowledge about alterations and functional roles of a large part of the identified up-regulated miRNAs is still missing, functional roles have been described for some of them. Remarkably, miR-654-5p (2.30-fold increase by Res) was recently found to be among miRs that target an extended AR 3' UTR and downregulate AR, prostate-specific antigen (PSA) and related androgen-induced proliferation in LNCaP cells [71]. Our own data on Res down-regulation of AR and PSA in LNCaP cells suggest a possible link between Res-up-regulated miR-654-5p and Res inhibition of AR and cell proliferation (unpublished data). Loss of miR-149 has been reported to cause gain of function of KCNMA1 and lysyl oxidase (LOX) oncogenes in clear cell renal cell carcinoma [72]. Notably, miR-149 was down-regulated in PCa, and was part of a sixmember diagnostic marker set that discriminated between malignant and normal tissues in PCa patients [29]. Downregulation of miR-150 in relapsed patients was associated with the prognosis and disease progression of pediatric acute leukemia [73]. Down-regulation of miR-150 and its tumor suppressor activity has been demonstrated in mouse model of B-cell lymphoma [44]. Moreover, miR-150, one of the main regulatory miRNAs of immune function, was found to be significantly down-regulated in peripheral blood and leukocytes of sepsis patients compared to healthy controls. This down-regulation was associated with increased levels of antiinflammatory cytokine IL-18 [74]. On the other hand, miR-150 is shown to promote gastric cell proliferation [75]. Further, reduced expression of miR-152 has been documented in nasopharyngeal carcinoma [76]. miR-152 was suggested as a tumor suppressor because very low levels were found in highly invasive melanoma cell line [77], and there was a reverse correlation with high levels of micropthalmiaassociated transcription factor (MITF), an oncogene involved in melanoma progression [78]. In addition, hypermethylation of miR-152 was found in 34-86% of primary breast cancer tissues [79]. The down-regulation of miR-575, miR-483 and miR-654 was reported in meningiomas [80] and reduced expression of miR-939 was found in tumors of long-surviving neuroblastoma patients [81]; however, the role of these miRNAs remains unknown. Our results show Res up-regulation of a group of miRNAs that were reported to be upregulated in ovarian cancer tissues (miR-572) [82], squamous cell carcinoma (miR-762) [83] and breast cancer (miR-1246) [84]. miR-513a-5p was up-regulated in human bronchial epithelial cells after exposure to diesel exhaust particles [85], and was reported to repress interferon-γ-induced apoptosis [86]. miR-612 was recently mentioned as one of the six putative miRNAs negatively regulating tumor suppressor p73 [87]. Curiously, up-regulation of miR-1224 was reported in diet-induced fatty liver models in which mice were fed with alcoholic and nonalcoholic diets [88]. To date, there is no information about the alteration/function of miR-1308 and miR-874 in cancer. Taken together, Res up-regulated tumor suppressor miRs that are lost or down-regulated in various cancers, including PCa.

In summary, the finding that Res down-regulates oncomiRs and up-regulates tumor suppressor miRNAs corroborates our general idea of Res as a powerful negative modulator of miRNA-associated tumorigenesis.

3.2 Validation of selected Res-regulated miRNAs by qRT-PCR and Western blot analysis

To validate our array data, we selected several miRNAs that were down- or up-regulated by Res in LNCaP cells and

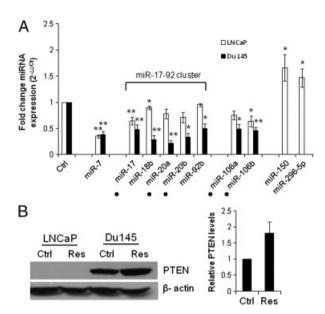


Figure 2. (A) Validation of selected miRNAs by qRT-PCR. LNCaP and Du145 cells were maintained in phenol red-free media/ stripped serum overnight prior to treatment with 50 µM Res. RNA was isolated and gRT-PCR was performed as described in Section 2. All samples were tested in duplicate, and experiments were repeated at least four times. All samples were within the measurable range (C_t <35). ΔC_t for each miR was calculated by subtracting the Ct of reference SNORD44 from the Ct of given miR. $\Delta\Delta C_t$ was then calculated by subtracting the ΔC_t of the EtOH-treated Ctrl from ΔC_t of the Res-treated samples. Fold changes for the expression of miRs were calculated using the equation $2^{-\Delta\Delta Ct}.$ Data are presented as the mean $\pm\,\text{SE}$ of four or more independent experiments. *p<0.05; **p<0.01 (Student's t-test); •, indicate miRs that target PTEN according to www. microRNA.org (miRanda) (B) Res up-regulates PTEN in Du145 cells. Left, LNCaP and Du145 cells were treated with 50 µM Res and EtOH (Ctrl) when in phenol red-free media with stripped serum. PTEN expression was detected by Western blot using whole cell extracts, and levels of PTEN (55 kDa) was detected. βactin was used as a loading control. Representative blot is shown. Right, quantification of PTEN protein levels in Du145 cells normalized to β -actin from three independent experiments: Res-treated cells express in average 1.8-fold higher PTEN than Ctrl cells. Bars, SE.

performed qRT-PCR (Fig. 2A). In agreement with the array data, changes by Res in each miR showed the same trend. Specifically, miR-7, miR-17, miR-18b, miR-20a, miR-92b, miR-106a and miR106b each decreased whereas miR-150 and miR-296-5p increased in Res-treated cells. The miRs from the miR-17-92 and miRs-106a and b oncogenic clusters were of particular interest because (i) of their importance for PCa tumorigenesis [26, 49, 51] and (ii) of their down-regulation by Res. Importantly, these two clusters are regulators of tumor suppressor PTEN, which is frequently defective in PCa. Since LNCaP cells are PTEN-negative [89, 90] and Du145 cells express wild-type PTEN [91] we pursued our

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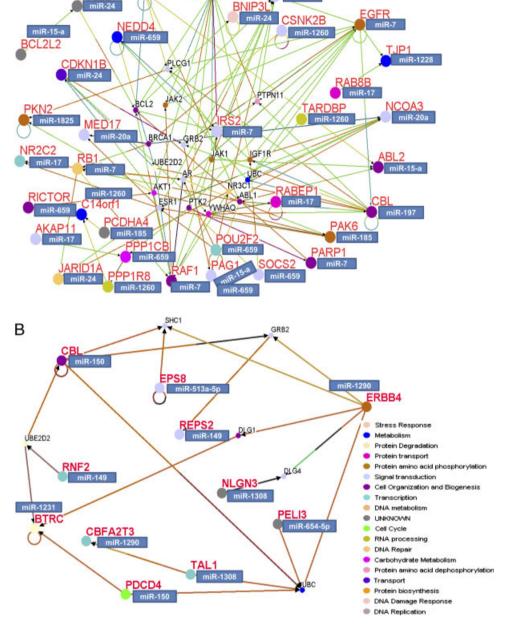


Figure 3. Interaction maps of miRNA potential targets (based in silico prediction). (A) Gene network of oncomiR target group. (B) Gene network of tumor suppressor miRNA target group. Genes are represented by circles, red letters. The "big" circles indicate miRrelated targets. Corresponding miRs are represented by boxes. The "small" circles indicate the intermediate genes from GO database, black letters. Lines represent interactions. Genes are color-coded according to class/function.

findings with Du145 cells and checked whether Res down-regulates miR-17-92 and -106a, b clusters in these cells. We found that Res significantly down-regulated miRs associated with PTEN regulation in Du145 cells (Fig. 2A). To find out whether Res up-regulates PTEN protein in PCa cells, we performed Western blot analysis and showed that Res up-regulated PTEN by about twofold in Du145 cells (Fig. 2B). Whether this up-regulation is a direct consequence of miRNA-related alterations still has to be established. LNCaP cells were PTEN-negative as has been previously reported [89, 90].

3.3 In silico network analysis of Res-regulated miRNA putative targets

It is established that multiple co-expressed miRNAs may work collectively to regulate related targets in common pathways [92]. To link Res-regulated miRNA profiling data with biological consequences, we took a computational approach to examine the possible pathways collectively regulated by the co-expressed down- or up-regulated miRNAs on the basis of their predicted targets. By applying a strict criteria of ≤ -0.5 context score, 308 targets were

identified in down-regulated (oncogenic) set, and 193 mRNAs in up-regulated (tumor suppressor) set. miRs that did not pass the threshold of context score and showed no predicted targets (Table 1) were excluded from further analyses.

We identified a potential gene network, i.e. the interaction between potential targets for each set of Res-regulated miRNAs (Fig. 3). Within the oncomiR target group (Fig. 3A), 11 (miR-7, miR-17, miR-1228, miR-20a, miR-197, miR-15a, miR-185, miR-659, miR-1260, miR-24, miR-1825) out of 18 down-regulated miRs accounted for the 36 predicted targets that form the potential gene network, including "connectors" from the GO data base. Genes in "signal transduction", "cell organization and biogenesis", protein phosphorylation and metabolism form the top gene networks within this set. Within the tumor suppressor miRNA target group (Fig. 3B), seven (miR-1290, miR-654-5p, miR-1308, miR-150, miR-1231, miR-149 and miR-513a-5p) out of 16 up-regulated miRs accounted for the 11 predicted targets in the "gene transcription" network.

Our results revealed that the targets co-regulated by the Res-down-regulated oncomiRs include several key regulators of carcinogenesis such as tumor suppressor gene retinoblastoma (RB), which controls androgen signaling and PCa progression [93], tumor suppressor poly (ADP-ribose) polymerase 1 (PARP1) [94], negative growth regulators (SOCS) [57, 58] and activators of pro-apoptotic pathways (BLC2L11) [95]. Interestingly, it has been shown recently that Res induces growth arrest and apoptosis in PCa cells through activation of forkhead box O (Foxo)-mediated up-regulation of BLC2L11), also known as Bim [96]. It is very possible that Res up-regulation of Bim in PCa might occur in part through down-regulation of miR-24. In contrast, the targets co-regulated by the Res-up-regulated tumor suppressors include oncogenes, activators of survival pathways and inhibitors of apoptosis. For example, REPS2 (RALBP1-associated Eps domain containing 2) inhibits epidermal growth factor (EGF) signaling in advanced metastatic PCa [97] and plays a role in overcoming resistance to apoptosis in androgen-resistant PCa [98], while reactivation of v-erb erythroblastic leukemia viral oncogene homolog 4 (ERBB-4) results in apoptosis of cancer cells [99]. Further studies are warranted to provide experimental validation of the complex interconnection between Res-induced co-regulated miRNAs and their targets.

4 Concluding remarks

In summary, this study provides clear evidence that Resinduced modulation on miRNA expression in PCa may contribute to the cancer-protective effects of Res. We found that Res down-regulated several important oncomiRs and up-regulated some tumor suppressor miRs in PCa cells, which potentially can lead to regulation of target mRNAs essential in carcinogenesis and cancer progression.

Although the functions of most miRNAs regulated by Res have yet to be discovered, it is apparent that Res-regulated miRNA profiles provide a basis for future experimental functional studies in vitro and in vivo to identify their physiological role in PCa and to offer a novel perspective on the potential chemopreventive and chemotherapeutic benefits of Res.

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The authors have declared no conflict of interest.

5 References

- [1] Stewart, J. R., Artime, M. C., O'Brian, C. A., Resveratrol: a candidate nutritional substance for prostate cancer prevention. J. Nutr. 2003, 133, 2440S–2443S.
- [2] Schoonen, W. M., Salinas, C. A., Kiemeney, L. A., Stanford, J. L., Alcohol consumption and risk of prostate cancer in middle-aged men. *Int. J. Cancer* 2005, *113*, 133–140.
- [3] Jang, M., Cai, L., Udeani, G. O., Slowing, K. V. et al., Cancer chemopreventive activity of resveratrol, a natural product derived from grapes. *Science* 1997, 275, 218–220.
- [4] Juan, M. E., Vinardell, M. P., Planas, J. M., The daily oral administration of high doses of trans-resveratrol to rats for 28 days is not harmful. J. Nutr. 2002, 132, 257–260.
- [5] Asensi, M., Medina, I., Ortega, A., Carretero, J. et al., Inhibition of cancer growth by resveratrol is related to its low bioavailability. Free Radic. Biol. Med. 2002, 33, 387–398.
- [6] Marier, J. F., Vachon, P., Gritsas, A., Zhang, J. et al., Metabolism and disposition of resveratrol in rats: extent of absorption, glucuronidation, and enterohepatic recirculation evidenced by a linked-rat model. *J. Pharmacol. Exp. Ther.* 2002, 302, 369–373.
- [7] Brakenhielm, E., Cao, R., Cao, Y., Suppression of angiogenesis, tumor growth, and wound healing by resveratrol, a natural compound in red wine and grapes. FASEB J. 2001, 15, 1798–1800.
- [8] Tseng, S. H., Lin, S. M., Chen, J. C., Su, Y. H. et al., Resveratrol suppresses the angiogenesis and tumor growth of gliomas in rats. Clin. Cancer Res. 2004, 10, 2190–2202.
- [9] Kozuki, Y., Miura, Y., Yagasaki, K., Resveratrol suppresses hepatoma cell invasion independently of its anti-proliferative action. *Cancer Lett.* 2001, 167, 151–156.
- [10] Kimura, Y., Okuda, H., Resveratrol isolated from Polygonum cuspidatum root prevents tumor growth and metastasis to lung and tumor-induced neovascularization in

- Lewis lung carcinoma-bearing mice. J. Nutr. 2001, 131, 1844–1849.
- [11] Link, A., Balaguer, F., Goel, A., Cancer chemoprevention by dietary polyphenols: promising role for epigenetics. *Biochem. Pharmacol.* 2010, 80, 1771–1792.
- [12] Li, L. C., Carroll, P. R., Dahiya, R., Epigenetic changes in prostate cancer: implication for diagnosis and treatment. J. Natl. Cancer Inst. 2005, 97, 103–115.
- [13] Narayanan, B. A., Narayanan, N. K., Re, G. G., Nixon, D. W., Differential expression of genes induced by resveratrol in LNCaP cells: P53-mediated molecular targets. *Int. J. Cancer*. 2003, 104, 204–212.
- [14] Beher, D., Wu, J., Cumine, S., Kim, K. W. et al., Resveratrol is not a direct activator of SIRT1 enzyme activity. *Chem. Biol. Drug Des.* 2009, 74, 619–624.
- [15] Borra, M. T., Smith, B. C., Denu, J. M., Mechanism of human SIRT1 activation by resveratrol. J. Biol. Chem. 2005, 280, 17187–17195.
- [16] Nishioka, K., Chuikov, S., Sarma, K., Erdjument-Bromage, H. et al., Set9, a novel histone H3 methyltransferase that facilitates transcription by precluding histone tail modifications required for heterochromatin formation. *Genes Dev.* 2002. 16, 479–489.
- [17] Xue, Y., Wong, J., Moreno, G. T., Young, M. K. et al., NURD, a novel complex with both ATP-dependent chromatinremodeling and histone deacetylase activities. *Mol. Cell.* 1998, 2, 851–861.
- [18] Kai, L., Wang, J., Ivanovic, M., Chung, Y. T. et al., Targeting prostate cancer angiogenesis through metastasis-associated protein 1 (MTA1). *Prostate* 2011, 71, 268–280.
- [19] Kai, L., Samuel, S. K., Levenson, A. S., Resveratrol enhances p53 acetylation and apoptosis in prostate cancer by inhibiting MTA1/NuRD complex. *Int. J. Cancer* 2009, *126*, 1538–1548.
- [20] Chuang, J. C., Jones, P. A., Epigenetics and microRNAs. Pediatr. Res. 2007, 61, 24R–29R.
- [21] Li, Y., Kong, D., Wang, Z., Sarkar, F. H., Regulation of microRNAs by natural agents: an emerging field in chemoprevention and chemotherapy research. *Pharm. Res.* 2010, 27, 1027–1041.
- [22] Saini, S., Majid, S., Dahiya, R., Diet, microRNAs and prostate cancer. Pharm. Res. 2010, 27, 1014–1026.
- [23] He, L., Hannon, G. J., MicroRNAs: small RNAs with a big role in gene regulation. *Nat. Rev. Genet.* 2004, 5, 522–531.
- [24] Ambros, V., microRNAs: tiny regulators with great potential. Cell 2001, 107, 823–826.
- [25] Zhang, B., Pan, X., Cobb, G. P., Anderson, T. A., microRNAs as oncogenes and tumor suppressors. *Dev. Biol.* 2007, 302, 1–12.
- [26] Volinia, S., Calin, G. A., Liu, C. G., Ambs, S. et al., A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc. Natl. Acad. Sci. USA* 2006, 103, 2257–2261.
- [27] Ozen, M., Creighton, C. J., Ozdemir, M., Ittmann, M., Widespread deregulation of microRNA expression in human prostate cancer. *Oncogene* 2008, 27, 1788–1793.

- [28] Lu, J., Getz, G., Miska, E. A., Alvarez-Saavedra, E. et al., MicroRNA expression profiles classify human cancers. *Nature* 2005, 435, 834–838.
- [29] Schaefer, A., Jung, M., Mollenkopf, H. J., Wagner, I. et al., Diagnostic and prognostic implications of microRNA profiling in prostate carcinoma. *Int. J. Cancer* 2010, *126*, 1166–1176.
- [30] Brase, J. C., Johannes, M., Schlomm, T., Falth, M. et al., Circulating miRNAs are correlated with tumor progression in prostate cancer. *Int. J. Cancer* 2011, 128, 608–616.
- [31] Athar, M., Back, J. H., Tang, X., Kim, K. H. et al., Resveratrol: a review of preclinical studies for human cancer prevention. *Toxicol. Appl. Pharmacol.* 2007, 224, 274–283.
- [32] Aggarwal, B. B., Bhardwaj, A., Aggarwal, R. S., Seeram, N. P. et al., Role of resveratrol in prevention and therapy of cancer: preclinical and clinical studies. *Anticancer Res.* 2004, 24, 2783–2840.
- [33] Lewis, B. P., Shih, I. H., Jones-Rhoades, M. W., Bartel, D. P. et al., Prediction of mammalian microRNA targets. *Cell* 2003, 115, 787–798.
- [34] Grimson, A., Farh, K. K., Johnston, W. K., Garrett-Engele, P. et al., MicroRNA targeting specificity in mammals: determinants beyond seed pairing. *Mol. Cell* 2007, 27, 91–105.
- [35] Livak, K. J., Schmittgen, T. D., Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods* 2001, 25, 402-408.
- [36] Levenson, A. S., Gehm, B. D., Pearce, S. T., Horiguchi, J. et al., Resveratrol acts as an estrogen receptor (ER) agonist in breast cancer cells stably transfected with ER alpha. *Int. J. Cancer* 2003, 104, 587–596.
- [37] Benitez, D. A., Pozo-Guisado, E., Alvarez-Barrientos, A., Fernandez-Salguero, P. M. et al., Mechanisms involved in resveratrol-induced apoptosis and cell cycle arrest in prostate cancer-derived cell lines. J. Androl. 2007, 28, 282–293
- [38] Kuwajerwala, N., Cifuentes, E., Gautam, S., Menon, M. et al., Resveratrol induces prostate cancer cell entry into s phase and inhibits DNA synthesis. *Cancer Res.* 2002, 62, 2488–2492.
- [39] Harper, C. E., Patel, B. B., Wang, J., Arabshahi, A. et al., Resveratrol suppresses prostate cancer progression in transgenic mice. *Carcinogenesis* 2007, 28, 1946–1953.
- [40] Seeni, A., Takahashi, S., Takeshita, K., Tang, M. et al., Suppression of prostate cancer growth by resveratrol in the transgenic rat for adenocarcinoma of prostate (TRAP) model. Asian Pac. J. Cancer Prev. 2008, 9, 7–14.
- [41] Ambs, S., Prueitt, R. L., Yi, M., Hudson, R. S. et al., Genomic profiling of microRNA and messenger RNA reveals deregulated microRNA expression in prostate cancer. *Cancer Res.* 2008, 68, 6162–6170.
- [42] Leite, K. R., Sousa-Canavez, J. M., Reis, S. T., Tomiyama, A. H. et al., Change in expression of miR-let7c, miR-100, and miR-218 from high grade localized prostate cancer to metastasis. *Urol. Oncol.* 2009, 29, 265–269.

- [43] Spahn, M., Kneitz, S., Scholz, C. J., Stenger, N. et al., Expression of microRNA-221 is progressively reduced in aggressive prostate cancer and metastasis and predicts clinical recurrence. *Int. J. Cancer* 2010, 127, 394–403.
- [44] Chang, T. C., Yu, D., Lee, Y. S., Wentzel, E. A. et al., Wide-spread microRNA repression by Myc contributes to tumorigenesis. *Nat. Genet.* 2008, 40, 43–50.
- [45] Hayashita, Y., Osada, H., Tatematsu, Y., Yamada, H. et al., A polycistronic microRNA cluster, miR-17-92, is overexpressed in human lung cancers and enhances cell proliferation. *Cancer Res.* 2005, 65, 9628–9632.
- [46] Takakura, S., Mitsutake, N., Nakashima, M., Namba, H. et al., Oncogenic role of miR-17-92 cluster in anaplastic thyroid cancer cells. *Cancer Sci.* 2008, 99, 1147–1154.
- [47] O'Donnell, K. A., Wentzel, E. A., Zeller, K. I., Dang, C. V. et al., c-Myc-regulated microRNAs modulate E2F1 expression. *Nature* 2005, 435, 839–843.
- [48] Dews, M., Homayouni, A., Yu, D., Murphy, D. et al., Augmentation of tumor angiogenesis by a Myc-activated microRNA cluster. *Nat. Genet.* 2006, 38, 1060–1065.
- [49] Sylvestre, Y., De Guire, V., Querido, E., Mukhopadhyay, U. K. et al., An E2F/miR-20a autoregulatory feedback loop. J. Biol. Chem. 2007, 282, 2135–2143.
- [50] Libertini, S. J., Tepper, C. G., Guadalupe, M., Lu, Y. et al., E2F1 expression in LNCaP prostate cancer cells deregulates androgen dependent growth, suppresses differentiation, and enhances apoptosis. *Prostate* 2006, 66, 70–81.
- [51] Poliseno, L., Salmena, L., Riccardi, L., Fornari, A. et al., Identification of the miR-106b~25 microRNA cluster as a proto-oncogenic PTEN-targeting intron that cooperates with its host gene MCM7 in transformation. Sci. Signal. 2010, 3, ra29.
- [52] Dahia, P. L., PTEN, a unique tumor suppressor gene. Endocr. Relat. Cancer 2000, 7, 115–129.
- [53] Suzuki, H., Freije, D., Nusskern, D. R., Okami, K. et al., Interfocal heterogeneity of PTEN/MMAC1 gene alterations in multiple metastatic prostate cancer tissues. *Cancer Res.* 1998, 58, 204–209.
- [54] Guled, M., Lahti, L., Lindholm, P. M., Salmenkivi, K. et al., CDKN2A, NF2, and JUN are dysregulated among other genes by miRNAs in malignant mesothelioma -A miRNA microarray analysis. *Gene. Chromosome. Canc.* 2009, 48, 615–623.
- [55] Dyrskjot, L., Ostenfeld, M. S., Bramsen, J. B., Silahtaroglu, A. N. et al., Genomic profiling of microRNAs in bladder cancer: miR-129 is associated with poor outcome and promotes cell death in vitro. Cancer Res. 2009, 69, 4851–4860.
- [56] Huang, Y. W., Liu, J. C., Deatherage, D. E., Luo, J. et al., Epigenetic repression of microRNA-129-2 leads to overexpression of SOX4 oncogene in endometrial cancer. *Cancer Res.* 2009, 69, 9038–9046.
- [57] Neuwirt, H., Puhr, M., Santer, F. R., Susani, M. et al., Suppressor of cytokine signaling (SOCS)-1 is expressed in human prostate cancer and exerts growth-inhibitory function through down-regulation of cyclins and cyclin-dependent kinases. Am. J. Pathol. 2009, 174, 1921–1930.

- [58] Wang, G., Wang, J., Sadar, M. D., Crosstalk between the androgen receptor and beta-catenin in castrate-resistant prostate cancer. *Cancer Res.* 2008, 68, 9918–9927.
- [59] Foekens, J. A., Sieuwerts, A. M., Smid, M., Look, M. P. et al., Four miRNAs associated with aggressiveness of lymph node-negative, estrogen receptor-positive human breast cancer. Proc. Natl. Acad. Sci. USA 2008, 105, 13021–13026.
- [60] Chou, Y. T., Lin, H. H., Lien, Y. C., Wang, Y. H. et al., EGFR promotes lung tumorigenesis by activating miR-7 through a Ras/ERK/Myc pathway that targets the Ets2 transcriptional repressor ERF. Cancer Res. 2010, 70, 8822–8831.
- [61] Reddy, S. D., Ohshiro, K., Rayala, S. K., Kumar, R., Micro-RNA-7, a homeobox D10 target, inhibits p21-activated kinase 1 and regulates its functions. *Cancer Res.* 2008, 68, 8195–8200.
- [62] Kefas, B., Godlewski, J., Comeau, L., Li, Y. et al., microRNA-7 inhibits the epidermal growth factor receptor and the Akt pathway and is down-regulated in glioblastoma. *Cancer Res.* 2008, 68, 3566–3572.
- [63] Jiang, L., Liu, X., Chen, Z., Jin, Y. et al., MicroRNA-7 targets IGF1R (insulin-like growth factor 1 receptor) in tongue squamous cell carcinoma cells. *Biochem. J.* 2010, 432, 199–205.
- [64] Wang, B. D., Kline, C. L., Pastor, D. M., Olson, T. L. et al., Prostate apoptosis response protein 4 sensitizes human colon cancer cells to chemotherapeutic 5-FU through mediation of an NF kappaB and microRNA network. *Mol. Cancer* 2010, 9, 98.
- [65] Du, L., Schageman, J. J., Subauste, M. C., Saber, B. et al., miR-93, miR-98, and miR-197 regulate expression of tumor suppressor gene FUS1. *Mol. Cancer Res.* 2009, 7, 1234–1243.
- [66] Wang, X., Tang, S., Le, S. Y., Lu, R. et al., Aberrant expression of oncogenic and tumor-suppressive micro-RNAs in cervical cancer is required for cancer cell growth. *PLoS One* 2008, 3, e2557.
- [67] Qin, W., Shi, Y., Zhao, B., Yao, C. et al., miR-24 regulates apoptosis by targeting the open reading frame (ORF) region of FAF1 in cancer cells. *PLoS One* 2010, 5, e9429.
- [68] Takamizawa, J., Konishi, H., Yanagisawa, K., Tomida, S. et al., Reduced expression of the let-7 microRNAs in human lung cancers in association with shortened postoperative survival. *Cancer Res.* 2004, 64, 3753–3756.
- [69] Cimmino, A., Calin, G. A., Fabbri, M., Iorio, M. V. et al., miR-15 and miR-16 induce apoptosis by targeting BCL2. Proc. Natl. Acad. Sci. USA 2005, 102, 13944–13949.
- [70] Takahashi, Y., Forrest, A. R., Maeno, E., Hashimoto, T. et al., MiR-107 and MiR-185 can induce cell cycle arrest in human non small cell lung cancer cell lines. *PLoS One* 2009, 4, e6677.
- [71] Ostling, P., Leivonen, S. K., Aakula, A., Kohonen, P. et al., Systematic analysis of MicroRNAs targeting the androgen receptor in prostate cancer cells. *Cancer Res.* 2011, 71, 1956–1967.
- [72] Liu, H., Brannon, A. R., Reddy, A. R., Alexe, G. et al., Identifying mRNA targets of microRNA dysregulated in cancer:

- with application to clear cell renal cell carcinoma. *BMC Syst. Biol.* 2010, *4*, 51.
- [73] Zhang, H., Luo, X. Q., Zhang, P., Huang, L. B. et al., Micro-RNA patterns associated with clinical prognostic parameters and CNS relapse prediction in pediatric acute leukemia. *PLoS One* 2009, 4, e7826.
- [74] Vasilescu, C., Rossi, S., Shimizu, M., Tudor, S. et al., Micro-RNA fingerprints identify miR-150 as a plasma prognostic marker in patients with sepsis. *PLoS One* 2009, 4, e7405.
- [75] Wu, Q., Jin, H., Yang, Z., Luo, G. et al., MiR-150 promotes gastric cancer proliferation by negatively regulating the proapoptotic gene EGR2. *Biochem. Biophys. Res. Commun.* 2010, 392, 340–345.
- [76] Chen, H. C., Chen, G. H., Chen, Y. H., Liao, W. L. et al., MicroRNA deregulation and pathway alterations in nasopharyngeal carcinoma. *Br. J. Cancer* 2009, 100, 1002–1011.
- [77] Mueller, D. W., Rehli, M., Bosserhoff, A. K., miRNA expression profiling in melanocytes and melanoma cell lines reveals miRNAs associated with formation and progression of malignant melanoma. *J. Invest. Dermatol.* 2009, 129, 1740–1751.
- [78] Haflidadottir, B. S., Bergsteinsdottir, K., Praetorius, C., Steingrimsson, E., miR-148 regulates Mitf in melanoma cells. PLoS One 2010, 5, e11574.
- [79] Lehmann, U., Hasemeier, B., Christgen, M., Muller, M. et al., Epigenetic inactivation of microRNA gene hsa-mir-9-1 in human breast cancer. J. Pathol. 2008, 214, 17–24.
- [80] Saydam, O., Shen, Y., Wurdinger, T., Senol, O. et al., Downregulated microRNA-200a in meningiomas promotes tumor growth by reducing E-cadherin and activating the Wnt/beta-catenin signaling pathway. *Mol. Cell. Biol.* 2009, 29, 5923–5940.
- [81] Scaruffi, P., Stigliani, S., Moretti, S., Coco, S. et al., Transcribed-Ultra Conserved Region expression is associated with outcome in high-risk neuroblastoma. BMC Cancer 2009. 9. 441.
- [82] Dahiya, N., Sherman-Baust, C. A., Wang, T. L., Davidson, B. et al., MicroRNA expression and identification of putative miRNA targets in ovarian cancer. PLoS One 2008, 3, e2436.
- [83] Yu, T., Wang, X. Y., Gong, R. G., Li, A. et al., The expression profile of microRNAs in a model of 7, 12-dimethylbenz[a]anthrance-induced oral carcinogenesis in Syrian hamster. J. Exp. Clin. Cancer Res. 2009, 28, 64.
- [84] Pigati, L., Yaddanapudi, S. C., Iyengar, R., Kim, D. J. et al., Selective release of microRNA species from normal and malignant mammary epithelial cells. *PLoS One* 2010, 5, e13515.
- [85] Jardim, M. J., Fry, R. C., Jaspers, I., Dailey, L. et al., Disruption of microRNA expression in human airway cells by diesel exhaust particles is linked to tumorigenesisassociated pathways. *Environ. Health Perspect.* 2009, 117, 1745–1751.
- [86] Gong, A. Y., Zhou, R., Hu, G., Li, X. et al., MicroRNA-513 regulates B7-H1 translation and is involved in IFN-gamma-

- induced B7-H1 expression in cholangiocytes. *J. Immunol.* 2009, *182*, 1325–1333.
- [87] Boominathan, L., The tumor suppressors p53, p63, and p73 are regulators of microRNA processing complex. PLoS One 2010, 5, e10615.
- [88] Dolganiuc, A., Petrasek, J., Kodys, K., Catalano, D. et al., MicroRNA expression profile in Lieber-DeCarli diet-induced alcoholic and methionine choline deficient diet-induced nonalcoholic steatohepatitis models in mice. *Alcohol Clin.* Exp. Res. 2009, 33, 1704–1710.
- [89] Davies, M. A., Koul, D., Dhesi, H., Berman, R. et al., Regulation of Akt/PKB activity, cellular growth, and apoptosis in prostate carcinoma cells by MMAC/PTEN. Cancer Res. 1999, 59, 2551–2556.
- [90] Myers, M. P., Pass, I., Batty, I. H., Van der Kaay, J. et al., The lipid phosphatase activity of PTEN is critical for its tumor supressor function. *Proc. Natl. Acad. Sci. USA* 1998, 95, 13513–13518.
- [91] Grunwald, V., DeGraffenried, L., Russel, D., Friedrichs, W. E. et al., Inhibitors of mTOR reverse doxorubicin resistance conferred by PTEN status in prostate cancer cells. *Cancer Res.* 2002, 62, 6141–6145.
- [92] Cloonan, N., Brown, M. K., Steptoe, A. L., Wani, S. et al., The miR-17-5p microRNA is a key regulator of the G1/S phase cell cycle transition. *Genome Biol.* 2008, 9, R127.
- [93] Sharma, A., Yeow, W. S., Ertel, A., Coleman, I. et al., The retinoblastoma tumor suppressor controls androgen signaling and human prostate cancer progression. J. Clin. Invest. 2010, 120, 4478–4492.
- [94] Wesierska-Gadek, J., Ranftler, C., Schmid, G., Physiological ageing: role of p53 and PARP-1 tumor suppressors in the regulation of terminal senescence. J. Physiol. Pharmacol. 2005, 56, 77–88.
- [95] Hoshino, Y., Katsuno, Y., Ehata, S., Miyazono, K., Autocrine TGF-beta protects breast cancer cells from apoptosis through reduction of BH3-only protein, Bim. J. Biochem. 2011, 149, 55–65.
- [96] Chen, Q., Ganapathy, S., Singh, K.P., Shankar, S. et al., Resveratrol induces growth arrest and apoptosis through activation of FOXO transcription factors in prostate cancer cells. PLoS One 2010, 5, e15288.
- [97] Oosterhoff, J. K., Kuhne, L. C., Grootegoed, J. A., Blok, L. J., EGF signalling in prostate cancer cell lines is inhibited by a high expression level of the endocytosis protein REPS2. *Int. J. Cancer* 2005, *113*, 561–567.
- [98] Penninkhof, F., Grootegoed, J. A., Blok, L. J., Identification of REPS2 as a putative modulator of NF-kappaB activity in prostate cancer cells. *Oncogene* 2004, 23, 5607–5615.
- [99] Das, P. M., Thor, A. D., Edgerton, S. M., Barry, S. K. et al., Reactivation of epigenetically silenced HER4/ERBB4 results in apoptosis of breast tumor cells. *Oncogene* 2010, 29, 5214–5219.